

- Jencks, W. P. (1980a) *Adv. Enzymol. Relat. Areas Mol. Biol.* 51, 75–106.
- Jencks, W. P. (1980b) in *Molecular Biology, Biochemistry and Biophysics, Chemical Recognition in Biology* (Chapeville, F., & Haenni, A. L., Eds.) Vol. 32, pp 11–12, Springer-Verlag, Berlin.
- Khananshvil, D., & Jencks, W. P. (1988) *Biochemistry* 27, 2943–2952.
- Klemens, M. R., & Grisham, C. M. (1988) *FEBS Lett.* 237, 4–8.
- Lowry, O. H., Rosebrough, A. L., Farr, A. L., & Randall, R. J. (1951) *J. Biol. Chem.* 193, 265–275.
- MacLennan, D. H., Brandl, C. J., Korczak, B., & Green, N. M. (1985) *Nature* 316, 696–700.
- Masuda, H., & deMeis, L. (1973) *Biochemistry* 12, 4581–4585.
- Morrison, J. F., & Cleland, W. W. (1983) *Biochemistry* 22, 5507–5513.
- Pauling, L. (1946) *Chem. Eng. News* 24, 1375–1380.
- Petithory, J. R., & Jencks, W. P. (1986) *Biochemistry* 25, 4493–4497.
- Petithory, J. R., & Jencks, W. P. (1988) *Biochemistry* 27, 5553–5564.
- Pickart, C. M., & Jencks, W. P. (1982) *J. Biol. Chem.* 257, 5319–5322.
- Pickart, C. M., & Jencks, W. P. (1984) *J. Biol. Chem.* 259, 1629–1643.
- Rossi, B., Leone, F. d. A., Gache, C., & Lazdunski, M. (1979) *J. Biol. Chem.* 254, 2302–2307.
- Shigekawa, M., & Dougherty, J. P. (1978) *J. Biol. Chem.* 253, 1451–1457.
- Shigekawa, M., Wakabayashi, S., & Nakamura, H. (1983) *J. Biol. Chem.* 258, 8698–8707.
- Stahl, N., & Jencks, W. P. (1984) *Biochemistry* 23, 5389–5392.
- Stahl, N., & Jencks, W. P. (1987) *Biochemistry* 26, 7654–7667.
- Sumida, M., Wang, T., Schwartz, A., Younkin, C., & Froehlich, J. P. (1980) *J. Biol. Chem.* 255, 1497–1503.
- Takakuwa, Y., & Kanazawa, T. (1979) *Biochem. Biophys. Res. Commun.* 88, 1209–1216.
- Takisawa, H., & Tonomura, Y. (1978) *J. Biochem. (Tokyo)* 83, 1275–1284.
- Verjovski-Almeida, S., Kurzmack, S., & Inesi, G. (1978) *Biochemistry* 17, 5006–5013.
- Vianna, A. L. (1975) *Biochim. Biophys. Acta* 410, 389–406.
- Wakabayashi, S., & Shigekawa, M. (1987) *J. Biol. Chem.* 262, 11524–11531.
- Wang, T. (1986) *J. Biol. Chem.* 261, 6307–6316.
- Yamada, S., & Tonomura, Y. (1972) *J. Biochem. (Tokyo)* 72, 417–425.
- Yamada, S., & Ikemoto, N. (1980) *J. Biol. Chem.* 255, 3018–3119.

CORRECTIONS

Iron(II)-Ethylenediaminetetraacetic Acid Catalyzed Cleavage of DNA Is Highly Specific for Duplex DNA, by Maria J. Jezewska, Włodzimierz Bujalowski, and Timothy M. Lohman*, Volume 28, Number 15, July 25, 1989, pages 6161–6164.

We have repeated a study of the Fe(II)-EDTA-catalyzed cleavage of single-stranded (ss) vs duplex DNA that was recently reported in the above mentioned paper in order to obtain a quantitative estimate of the relative preference for cleavage of duplex DNA. Upon repeating these studies, we did *not* observe the dramatic difference in cleavage of ss vs duplex DNA that we had previously reported. We do still observe that the duplex [d(pT)₇₀·dA(pA)₆₉] is cleaved preferentially compared to d(pT)₇₀ (pH 7.2); however, this preference is *only moderate*, rather than highly specific as we originally reported. The disappearance of the ³²P-labeled intact d(pT)₇₀ when single stranded vs when in a duplex with dA(pA)₆₉ was monitored after separation of the reaction products by polyacrylamide gel electrophoresis (in 7 M urea). The amount of ³²P-labeled d(pT)₇₀ was quantitated with a Betascope 603 blot analyzer (Betagen, Waltham, MA). The cleavage reaction was performed as a function of the concentration of Fe(II)-EDTA (20, 50, 100, and 200 μM) as described previously. The results indicate only a slight preference (factor of 1.3–2) for cleavage

of duplex DNA, based on six repetitions of this experiment. At this time we cannot explain the basis for the different quantitative results that we now observe vs those reported previously. On the basis of these recent data, we conclude that the cleavage of DNA that is catalyzed by Fe(II)-EDTA shows only a modest preference for duplex over single-stranded nucleic acids, which is in agreement with the recent report by Celander and Cech [Celander, D. W., & Cech, T. R. (1990) *Biochemistry* 29, 1355–1361], who also reported only a slight preference for cleavage of duplex over single-stranded DNA and RNA. In any event, this small preference for cleavage of duplex DNA indicates that the mechanism of the cleavage reaction requires further study.

Glycinamide Ribonucleotide Synthetase from *Escherichia coli*: Cloning, Overproduction, Sequencing, Isolation, and Characterization, by Y. S. Cheng, J. Rudolph, M. Stern, J. Stubbe,* K. A. Flannigan, and J. M. Smith*, Volume 29, Number 1, January 9, 1990, pages 218–227.

Page 218. The first author on this paper should be Y. S. Cheng and *not* Y. Shen.